IN THE SPECIFICATION

Please amend the paragraph at page 9, lines 11 to 22 as follows:

In this invention the targeting protein, such as a hydrophobin-like protein or parts of it, is bound to the product molecule or the component to be separated. First, phase forming materials and eventually possibly also additional salts are added to a watery solution containing the fusion molecule or component, and optionally also the contaminating materials. The added agents are mixed to facilitate their solubilization. As soon as they are solubilized the two phase are formed either by gravity settling or centrifugation. In the separation the ~~target~~ targeting protein drives the product to for instance the detergent-rich phase which could either be the top or the bottom phase. The method is not only useful for purification of products of interest but also for keeping the product or the component of interest, such as a biocatalyst, in a particular phase which enables certain useful biotechnical reactions.

Please amend the paragraph at page 20, lines 18 to 26 as follows:

In general whole fermentation broth, supernatant (biomass separated by centrifugation or filtration) or purified proteins in buffer were separated in 10 ml graduated tubes. First detergent was added into the tubes and the tubes were then filled to 10 ~~mg~~ ml with protein containing liquid. The amount of detergent in the tube was calculated in weight percent of detergents. After thorough mixing in an overhead shaker the separation took place by either gravity settling in a water bath at constant temperature or by centrifugation at constant temperature. The separation usually was performed at 30°C, the standard amount of detergent used was 2-5% (w/v). After separation the volume ratio was noted and samples were taken from the lighter and heavier phase for analysis.

Please amend the paragraph at page 21, lines 24 to 31 as follows:

The partition coefficient K was defined as the ratio of the measured concentrations or activities in the top and bottom phase, respectively.

The Yield Y was defined as follows:

$$Y_T = \frac{1}{1+\left[\frac{V_B}{V_T}\cdot\frac{1}{K}\right]}$$

where Y_T is the Yield of the top phase, V_B and V_T are the volumes of ~~top and~~ bottom and top phase, respectively. The Yield of the bottom phase can be described accordingly.

Please amend the paragraph at page 29, lines 6 to 9, as follows:

The effect of HFBI on partitioning and the final yield can further be demonstrated by comparing the extraction result of ~~EGIcore-HFB~~ EGIcore-HFBI fusion with extraction results obtained with purified wild type EGI and EGIcore. The fusion protein is partitioning more than 100 times better to the detergent phase (see table ~~belove~~ below).

Please amend the paragraph at page 45, line 23 to page 46, line 3 as follows:

For preparative purposes, HFBI and ~~HFBI~~ HFBII were extracted from 0.5 l of culture supernantants from cultivations carried out on media containing glucose or cellulose as described in Example 4. 2% *(w/w)* detergent was added to the culture supernatant which was mixed and then allowed to settle in a separation funnel at 20°C if C11E02 (Berol 532) was used and 30°C if C12-18E05 (Henkel) was used. The detergent phase (enriched phase) was collected and mixed with an equal volume of isobutanol. When C11E02 was used an equal volume of 50 mM acetate buffer was also added. The remaining culture supernatant is the depleted phase. To follow the purification, analytical HPLC was run on samples from each step (Figure 27 and 28). On the basis of HPLC analysis, both HFBI and HFBII partitioned well to both of the detergents. When C11E02 was used, K values of >1000 and >78 were obtained for HFBI and HFBII, respectively.